

Diagnosis of *Strongyloides stercoralis* infection in immunocompromised patients: problems and perspectives

Diagnóstico da infecção por *Strongyloides stercoralis* em pacientes imunocomprometidos: problemas e perspectivas

Fabiana Martins de Paula¹, Ronaldo César Borges Gryscek², Pedro Paulo Chieffi³

Abstract

The main aspects and problems related to laboratory diagnosis of the human infection by *Strongyloides stercoralis*, especially in the case of immunocompromised patients, are discussed. The perspectives for improving the performance of diagnostic tests are also appointed.

Keywords: Strongyloidiasis, Clinical laboratory techniques, Patients, Immunocompromised hosts

Resumo

O presente artigo discute os principais aspectos e problemas relacionados ao diagnóstico laboratorial da infecção por *Strongyloides stercoralis* em pacientes imunocomprometidos. São abordadas, também, as perspectivas para aperfeiçoamento dos testes diagnósticos na estrogiloidíase humana.

Descritores: Estrogiloidíase, Técnicas de laboratório clínico, Pacientes, Hospedeiro imunocomprometido

Strongyloidiasis is a parasitic disease caused by the intestinal nematode *Strongyloides stercoralis*, prevalent

in several parts of the world, especially in tropical and subtropical regions such as Latin America. The evaluation of the human infection by *Strongyloides stercoralis* is not quite precise; according to some authors it is estimated that 30 to 370 million people are infected with this helminth throughout the world⁽¹⁻³⁾.

This helminthiasis, more often, is responsible for chronic asymptomatic or oligosymptomatic infections of the gastrointestinal tract in immunocompetent human hosts and may remain undiagnosed for long time⁽⁴⁻⁵⁾. However, in immunocompromised patients, this infection may be extremely serious, disclosing such as hyperinfection syndrome and / or disseminated disease with high morbidity and lethality⁽⁶⁻⁷⁾. Therefore, the diagnosis and early treatment of this helminthiasis is essential for a favorable prognosis⁽⁸⁻⁹⁾.

The definitive diagnosis of strongyloidiasis is performed mainly by the identification of larvae in faecal samples, using concentration techniques or culture methods^(5,10). However, these larvae are excreted irregularly and, sometimes, in a small number. So, the coproparasitological methods show low sensitivity for the detection of this parasite in many cases, requiring the analysis of several faecal samples to increase the sensibility of the test^(5,10-11). Considering the group of patients submitted to immunosuppression, in some occasions, the infection by *Strongyloides stercoralis* is not suspected, resulting in no request for coproparasitological diagnostic tests.

In contrast, several immunological tests, with increased sensitivity and specificity, have been proposed for the diagnosis of *Strongyloides stercoralis* human infection. The ELISA test has been highlighted mainly by the practicality of the analysis of several samples together, while Western blotting is indicated by the recognition of antigenic protein fractions of the parasite⁽¹²⁻¹⁴⁾. However, the use of these methods has some limitations, such as cross-reactivity with other helminthic infections, serological positivity resulting from previous *Strongyloides stercoralis* infections or, even, low sensitivity in immunocompromised patients. Another limitation for the immunodiagnosis of

1. Pesquisador Científico do Laboratório de Investigação Médica (LIM/06 - Laboratório de Imunopatologia da Esquistossomose). Hospital das Clínicas da Faculdade de Medicina da Universidade de São Paulo. São Paulo – SP – Brasil

2. Professor Associado da Universidade de São Paulo. Faculdade de Medicina. Departamento de Moléstias Infecciosas e Parasitárias. São Paulo – SP – Brasil

3. Professor Emérito da Faculdade de Ciências Médicas da Santa Casa de São Paulo. São Paulo – SP – Brasil

Institution: Universidade de São Paulo. Instituto de Medicina Tropical de São Paulo / Faculdade de Ciências Médicas da Santa Casa de São Paulo. Departamento de Ciências Patológicas. São Paulo – SP – Brasil

Correspondence Address: Prof. Dr. Pedro Paulo Chieffi. Faculdade de Ciências Médicas da Santa Casa de São Paulo. Departamento de Ciências Patológicas. Rua Dr. Cesário Mota Jr, 61 – Vila Buarque – 021021-020 – São Paulo – SP – Brasil

strongyloidiasis is the difficulty for obtaining enough amounts of *Strongyloides stercoralis* larvae or adults for production of specific antigens. This fact justifies the need for standardization and use of heterologous antigens, especially in the endemic areas^(12, 15-17). Thus, wild-rodent species *Strongyloides venezuelensis* and *Strongyloides ratti* have been employed for the development of immunological techniques for the study and diagnosis of human strongyloidiasis^(15,18).

On the other hand, the development and implementation of new studies involving proteomic techniques will be of fundamental importance for the accurate serological diagnosis of *Strongyloides stercoralis* infection in the future^(13-14,19-20). In this context, using the *Strongyloides venezuelensis* heterologous species as an alternative source for antigen production, proteins recognized by antibodies present in the sera of patients with strongyloidiasis has been evaluated⁽¹⁴⁾.

Trying to implement the diagnosis of strongyloidiasis, molecular tests have been used in recent years⁽²¹⁻²³⁾. Although using molecular biology techniques in the context of strongyloidiasis diagnosis present promising results, some problems must be considered: the target region for DNA amplification, the presence of inhibitors in the faecal material, the lack of a standard protocol for DNA extraction, and small amount of faecal material in collected samples are challenges that yet must be solved. These obstacles may compromise the sensitivity of the trials⁽²¹⁻²²⁾. However, the wide range of proteins secreted by *Strongyloides stercoralis* larvae and adults recently identified, that have been previously found in other helminth species, must be considered as an additional problem for the specificity of these diagnostic techniques⁽²⁴⁾. On the other hand, there are scarce number of reference laboratories that would provide the precise methodology⁽²⁵⁾, mainly constituting a distant reality in the endemic areas⁽²⁶⁾.

Although the scientific literature demonstrates the importance and applicability of serological and molecular diagnosis, similar to the parasitological diagnosis, these are not yet systematically used in the screening of immunocompromised patients, due to the unavailability of these techniques in routine laboratories.

References

- Siddiqui AA, Berk SL. Diagnosis of *Strongyloides stercoralis* infection. Clin Infect Dis. 2001; 33(7):1040-7.
- Marcos LA, Terashima A, Canales M, Gotuzzo E. Update on strongyloidiasis in the immunocompromised host. Curr Infect Dis Rep. 2011; 13(1):35-46.
- Albonico M, Becker SL, Odermatt P, Angheben A, Anselmi M, Amor A, et al. StrongNet: an international network to improve diagnostics and access to treatment for strongyloidiasis control. PLoS Negl Trop Dis. 2016; 10(9): e0004898.
- Concha R, Harrington W Jr, Rogers AI. Intestinal strongyloidiasis: recognition, management, and determinants of outcome. J Clin Gastroenterol. 2005; 39(3):203-11.
- Toledo R, Muñoz-Antoli C, Esteban JG. Strongyloidiasis with emphasis on human infections and its different clinical forms. Adv Parasitol. 2015; 88:165-241.
- Keiser PB, Nutman TB. *Strongyloides stercoralis* in the immunocompromised population. Clin Microbiol Rev. 2004;17(1):208-17.
- Mejia R, Nutman TB. Screening, prevention, and treatment for hyperinfection syndrome and disseminated infections caused by *Strongyloides stercoralis*. Curr Opin Infect Dis. 2012; 25(4):458-63.
- Olsen A, van Lieshout L, Marti H, Polderman T, Polman K, Steinmann P, et al. Strongyloidiasis--the most neglected of the neglected tropical diseases? Trans R Soc Trop Med Hyg. 2009; 103(10):967-72.
- Krolewiecki AJ, Ramanathan R, Fink V, McAuliffe I, Cajal SP, Won K, et al. Improved diagnosis of *Strongyloides stercoralis* using recombinant antigen-based serologies in a community-wide study in northern Argentina. Clin Vaccine Immunol. 2010; 17(10):1624-30.
- Levenhagen MA, Costa-Cruz JM. Update on immunologic and molecular diagnosis of human strongyloidiasis. Acta Trop. 2014; 135:33-43.
- Uparanukraw P, Phongsri S, Morakote N. Fluctuations of larval excretion in *Strongyloides stercoralis* infection. Am J Trop Med Hyg. 1999; 60(6):967-73.
- Feliciano ND, Gonzaga HT, Gonçalves-Pires MR, Gonçalves AL, Rodrigues RM, Ueta MT, et al. Hydrophobic fractions from *Strongyloides venezuelensis* for use in the human immunodiagnosis of strongyloidiasis. Diagn Microbiol Infect Dis. 2010; 67(2):153-61.
- Rodpai R, Intapan PM, Thanchomngang T, Sanpool O, Janwan P, Laummaunwai P, et al. *Strongyloides stercoralis* diagnostic polypeptides for human strongyloidiasis and their proteomic analysis. Parasitol Res. 2016; 115(10):4007-12.
- Corral MA, Paula FM, Meisel DM, Castilho VL, Gonçalves EM, Levy D, et al. Potential immunological markers for diagnosis of human strongyloidiasis using heterologous antigens. Parasitology. 2017; 144(2):124-30.
- Costa-Cruz JM, Bullamah CB, Gonçalves-Pires MR, Campos DM, Vieira MA. Cryo-microtome sections of coproculture larvae of *Strongyloides stercoralis* and *Strongyloides ratti* as antigen sources for the immunodiagnosis of human strongyloidiasis. Rev Inst Med Trop Sao Paulo. 1997; 39(6):313-7.
- Corral MA, Paula FM, Gottardi M, Meisel DM, Chieffi PP, Gryscek RC. Membrane fractions from *Strongyloides venezuelensis* in the immunodiagnosis of human strongyloidiasis. Rev Inst Med Trop Sao Paulo. 2015; 57(1):77-80.
- Eamudomkarn C, Sithithaworn P, Sithithaworn J, Kaewkes S, Sripa B, Itoh M. Comparative evaluation of *Strongyloides ratti* and *S. stercoralis* larval antigen for diagnosis of strongyloidiasis in an endemic area of opisthorchiasis. Parasitol Res. 2015;114(7):2543-51.
- Bosqui LR, Gonçalves AL, Gonçalves-Pires MR, Custodio LA, de Menezes MC, Murad VA, et al. Detection of parasitespecific IgG and IgA in paired serum and saliva samples for diagnosis of human strongyloidiasis in northern Paraná state, Brazil. Acta Trop. 2015; 150:190-5.
- Marcilla A1, Sotillo J, Pérez-García A, Igual-Adell R, Valero ML, Sánchez-Pino MM, et al. Proteomic analysis of *Strongyloides stercoralis* L3 larvae. Parasitology, 2010; 137(10):1577-83.
- Soblik H, Younis AE, Mitreva M, Renard BY, Kirchner M, Geisinger F, et al. Life cycle stage-resolved proteomic analysis of the excretome/secretome from *Strongyloides ratti* identification of stage-specific proteases. Mol Cell Proteomics. 2011; 10(12):M111.010157.

21. Verweij JJ, Canales M, Polman K, Ziem J, Brienen EA, Polderman AM, et al. Molecular diagnosis of *Strongyloides stercoralis* in faecal samples using real-time PCR. *Trans R Soc Trop Med Hyg.* 2009; 103(4):342-6.
22. Repetto SA, Alba Soto CD, Cazorla SI, Tayeldin ML, Cuello S, Lasala MB, et al. An improved DNA isolation technique for PCR detection of *Strongyloides stercoralis* in stool samples. *Acta Trop.* 2013; 126(2):110-4.
23. Sitta RB, Malta FM, Pinho JR, Chieffi PP, Gryscek RC, Paula FM. Conventional PCR for molecular diagnosis of human strongyloidiasis. *Parasitology.* 2014; 141(5):716-21.
24. Maeda Y, Palomares-Rius JE, Hino A, Afrin T, Mondal SI, Nakatake A, et al. Secretome analysis of *Strongyloides venezuelensis* parasitic atages reveals that soluble and insoluble proteins are involved in its parasitism. *Parasit Vectors.* 2019; 12:21.
25. Bounfrate D; Formenti F, Perandin F, Bisoffi Z. Novel approaches to the diagnosis of *Strongyloides stercoralis* infection. *Clin Microbiol Infect.* 2015;21(6):543-52.
26. Paula FM, Malta FM, Marques PD, Sitta RB, Pinho JR, Gryscek RC, et al. Molecular diagnosis of strongyloidiasis in tropical areas: a comparison of conventional and real-time polymerase chain reaction with parasitological methods. *Mem Inst Oswaldo Cruz.* 2015; 110(2):272-4.

Trabalho recebido: 12/08/2019

Trabalho aprovado: 20/08/2019

Trabalho publicado: 20/08/2019